

Ocular Application of Nerve Growth Factor Protects Degenerating Retinal Ganglion Cells in a Rat Model of Glaucoma

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Purpose: Elevated intraocular pressure is a crucial pathologic event for the development of glaucoma (GL). We have reported that nerve growth factor (NGF) reaches retinal cells and the optic nerve (ON) when applied to the eye. Whether ocular application of NGF prevents or reduces damage to retinal ganglion cell (RGC) is not known.

Methods: GL was induced in adult rats by the injection of hypertonic saline into the episcleral vein of the right eye and the left eye used as control. Rats were then treated daily with ocular application of NGF or vehicle solution for 7 weeks. Retinal and ON tissues were then used for structural, immunohistochemical, and biochemical studies.

Results: The injection of hypertonic saline into the episcleral vein led to progressive degeneration of RGCs, with the loss of nearly 40% of these cells after 7 weeks of treatment. This cellular loss is associated with the downregulation of NGF and NGF-receptor expression in the retina and ON of the glaucomatous eye and ocular treatment with NGF significantly reduced the deficit induced by GL.

Conclusions: These findings indicate that NGF can exert protective action on RGC degeneration occurring in glaucomatous retina. We suggest that ocular NGF treatment might be a suitable pharmacologic approach to investigate protective mechanisms of degenerating RGCs.

Key Words: glaucoma (GL), intraocular pressure (IOP), NGF-receptor, neurotrophins, retinal ganglion cell (RGC)

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Glaucoma (GL) is the second leading cause of blindness worldwide and the prevalence of this disease is expected to grow as the population ages.¹ GL is a disease

characterized by progressive death of the retinal ganglion cells (RGCs) leads to optic nerve (ON) degeneration and vision loss. However, although the elevated intraocular pressure (EIOP) is considered a primary cause of the visual deficit, it is known that some patients still experience visual loss after lowering the IOP and others with GL and ON degeneration can have normal IOP.^{2–4} Despite these evidence, a major therapeutic aim is to facilitate the survival of RGCs through pharmacologic or surgical reduction of IOP.^{5–8} These and other observations have led to the hypothesis that degeneration of retinal cells may result from a secondary insult induced by alterations in the neuronal environment, such as changes in neurotransmitters, influx of calcium into the cells, formation of free radicals, or depletion of neurotrophic factors.^{5–8}

Nerve growth factor (NGF) is the first identified and best-characterized neuroprotective molecule, acting upon and produced by a number of cells within and outside the nervous system.^{9–11} The biologic activity of NGF is regulated by 2 different types of receptors that are located on the surface of responsive cells: the high-affinity NGF-receptor (TrkA), which belongs to the family of tyrosine kinase receptors, and the low-affinity NGF-receptor (p75), which is a transmembrane glycoprotein that lacks a tyrosine kinase domain.^{12,13} There is now a number of studies demonstrating that NGF is able to exert a wide spectrum of effects also on cells of the visual system, playing a crucial role in promoting the survival and growth of retinal cells. Thus, abnormal presence of NGF or its receptors can lead to cell death, not only in the central and peripheral nervous system,^{9–11} but also in the visual system *in vivo*^{14–22} and *in vitro*.^{23,24} NGF promotes the functional recovery of RGCs in an animal model of ocular ischemia,^{16–21} reduces retinal cell damage induced by intraocular hypertension,¹⁹ and delays retinal cell degeneration in rodents with retinitis pigmentosa.¹⁷ The functional role of NGF on retinal cells is supported by studies showing that intravitreal injection of NGF increases RGC survival and that eye NGF deprivation is known to lead to RGC death.¹⁹ One primary obstacle to clinical testing of NGF for retinal disorders is, however, the lack of an efficient, noninvasive means to deliver this factor in the posterior segment of the eye to the target cells. We have recently reported that intraocular administration of radiolabeled NGF binds to RGCs and ON axons²⁵ and that conjunctival applied NGF reaches retinal cells and brain NGF-receptive neurons.^{26,27} These observations suggested to investigate whether eye application of NGF protects retinal cells degeneration in a rat model of GL induced by EIOP in adult rats. The results of

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these studies indicate that GL induced in rats with EIOP reduces significantly the presence of NGF and the number of RGCs and ON axons, whereas ocular NGF administration significantly protects these degenerative events.

METHODS

Animals and Surgical Procedures

GL was induced in pathogen-free adult male Sprague-Dawley rats ($n = 94$), after anesthesia (ketamine 100 mg/mL; xylazine 20 mg/mL; and acepromazine 10 mg/mL). Briefly, as described by Morrison et al,²⁸ after a lateral canthotomy, a small plastic ring was fitted around the globe, straddling the equator and oriented to provide unobstructed passage for one radial aqueous vein in the superior quadrant of the eye, and then rats received 50 μ L sterile hypertonic saline solution (1.75 M NaCl) into the superior episcleral vein of the right eye. All animals were maintained in a 12 hours (06:00 to 18:00) light: dark cycle. The right eye was indicated as the glaucomatous eye and the left eye was indicated as control eye. Operated rats were divided into 2 groups and housed in single cages in a constant low-light environment (40 to 90 lx) to minimize IOP circadian oscillations. IOP was measured weekly with the TonoPen XL tonometer (Medtronic Ophthalmics, Jacksonville, FL) under topical anesthesia. IOP is reported as the mean of 10 valid readings per eye. Mean changes were calculated for each eye \pm SEM. The housing, care, and experimental procedures involving the experimental rats were carried out in accordance with the guidelines set by the Association for Research in Vision and Ophthalmology Statement for the Use of Animals in Ophthalmic and Vision Research and by local Ethical Commission on Animal Experimentation.

Animal Treatments

NGF was purchased from Biolab (Rome, Italy) or purified from adult male mouse submandibular glands following the method of Bocchini and Angeletti.²⁹ Purified NGF was dissolved in 0.9% NaCl phosphate (200 μ g/mL) and kept at 4°C for no longer than 1 week. Under these conditions, both stability and activity remained unchanged. Rats ($n = 42$) received topical ocular administration of 20 μ L vehicle solution, consisting of 0.9% NaCl, twice a day in both the right glaucomatous eye (GLV) and the left non-glaucomatous control eye (CV) for 7 weeks.

A second group of rats ($n = 42$) received topical ocular administration of 20 μ L of this NGF solution (4 μ g of purified NGF) 2 times a day for 7 weeks in the right glaucomatous eye (GLNGF) and the left control eye (CNGF). Animals were sacrificed with an overdose of Nembutal at time 0, 1, 3, and 7 weeks after the induction of GL.

Histologic Analysis of the Retina

For light histologic analysis of the retina, 5 eyes with attached ON were removed from 5 different animals of each experimental group ($n = 4$), fixed in Bouin fluid for 1 week, and then immersed in phosphate-buffered saline (PBS), pH 7.4, containing 20% sucrose for 3 days as described earlier.²⁶ Coded sections of the eyes were cut at 20 μ m with a cryostat (Leica CM 1850 UV, Germany) at -20°C , taking care that the cross sections of each retina were performed in the same orientation. Sections were then stained with hematoxylin-eosin. The number of RGCs were counted under a Zeiss microscope equipped with a

40 \times objective. To obtain representative data regarding the number of RGCs, RGC counts were performed on 5 random areas, located approximately 2 mm from the centre of the optic disc, of 6 different sections of each retina. All counts were performed in a masked manner and the results were averaged and converted to cells/mm².

Ultrastructural Analysis of the ON

The ON, located 1 mm from the adjacent retina, was dissected out, was fixed in 4% paraformaldehyde in 0.1 M PBS, pH 7.4, for 24 hours and then post-fixed in 1% OsO₄ for 4 hours, dehydrated with ascending ethanol, and then embedded in 812/Spurr's low viscosity resin. ON sections were cut with a Reichert microtome, collected on 200 mesh grids, stained with uranyl-acetate and lead citrate, observed under a Philips 100 electron microscope, and photographed. The presence of degenerating axons in 10 randomly selected fields, 50 \times 80 μ m, of each section ($n = 6$) of ON of each experimental group ($n = 4$), was examined and the number of axons with degenerating axon profiles, displaying myelin swelling and irregular debris, was counted in a masked manner and compared.

NGF-receptor Immunohistochemistry

Eyes were fixed overnight in 4% paraformaldehyde in 0.1 M PBS, pH 7.4, and then left overnight in 0.1 M PBS containing 20% sucrose. Coded 15- μ m thick sections of each retina were cut with a cryostat at -20°C and immunostained with anti-TrkA (1:70; Upstate, Temecula, CA) and anti-p75 antibodies, previously produced.²⁷ For quantitative analyses, we counted labeled RGCs in 12 different areas (10.0 \times 10²/mm² each) of each retina, located approximately 300 μ m from the centre of the optic disk, using a Zeiss microscope, equipped with a computerized image analysis system and a controlled motorized stage (IAS 2000, Delta Sistemi, Roma, Italy). The number of TrkA and p75 immunostained RGCs was defined as the percentage of immunostained cells compared with CV. All counts were performed in a masked manner.

Western Blotting

Retinal tissues were isolated and immediately sonicated. Tissue samples were homogenized in buffer (10 mM Tris-acetate, pH 7.4; 100 mM NaCl; 1 mM ethylenediamine-tetraacetic acid; 1 mM ethyleneglycol-tetraacetic acid; 1% Triton X-100; 10% glycerol; 0.1% sodium-dodecylphosphate, SDS; 20 mM Na₄P₂O₇; 2 mM sodium-orthovanadate, Na₃VO₄; 1 mM NaF; 2 μ g/mL aprotinin; 1 mM phenylmethylsulfonyl fluoride; and 1 μ g/mL leupeptin) at 4°C. After 12,000 rpm centrifugation for 20 minutes the supernatants were submitted to western blotting. Samples (30 μ g of total protein) were dissolved in loading buffer (0.1 M Tris-HCl buffer, pH 6.8, containing 0.2 M dithiothreitol, 4% SDS, 20% glycerol, and 0.1% bromophenol blue), separated by 8% or 12% SDS-polyacrylamide gel electrophoresis, and electrophoretically transferred to polyvinylidene difluoride membrane overnight. The membranes were incubated for 1 hour at room temperature with blocking buffer constituted by 5% non-fat dry milk in TBS-T (10 mM Tris, pH 7.5, 100 mM NaCl, and 0.1% Tween-20). Membranes were washed 3 times for 10 minutes each at room temperature in TBS-T followed by incubation at 4°C with primary antibodies overnight (anti-TrkA and anti p75 antibodies purchased from SantaCruz, CA and anti- β -actin from SIGMA, St Louis). Membranes were washed 3 times

for 10 minutes each at room temperature in TBS-T and incubated for 1 hour with horseradish peroxidase (HRP)-conjugated anti-rabbit IgG or HRP-conjugated anti-mouse IgG as the secondary antibody (Cell Signaling Technology, MA). The blots were developed with an enhanced chemiluminescent HRP substrate as the chromophore (Millipore, MA). The public Image J Software was used to evaluate band density, which was expressed as arbitrary units of grey level. The Image J program determines the optical density of the bands using a grey scale shareholding operation. The optical density of β -actin bands was used as a normalizing factor. For each gel per blot, the normalized values were then expressed as percentage of relative normalized controls and used for statistical evaluation. Statistical evaluations were performed using the StatView package for Windows. The data are expressed as means \pm SEM. TrkA and p75 protein concentrations determined by western blot analyses were evaluated by computing analyses of variance. A *P* value of less than 0.05 was considered significant.

Analysis of Cell Death

For the determination of RGC death and survival, sections of the retina were immunostained with terminal deoxynucleotidyl transferase-mediated deoxyuridine triphosphate-biotin nick-end labeling (TUNEL), a marker of apoptotic cell death.³⁰ Briefly, retinal sections were first incubated in a blocking solution (3% H₂O₂ in methanol) for 10 minutes at 15 to 25°C, incubated in a permeabilization solution containing PBS 1 \times with 0.1% Triton X-100 for 2 minutes at 4°C, and then labeled with an in-situ cell death detection kit (Roche Diagnostic, Boehringer, Mannheim, Germany) according to the manufacturer's instructions. DNA strand breaks were labeled and visualized with 0.4% diaminobenzidine-H₂O₂. TUNEL-positive cells with nuclear condensation or fragmentation were considered as apoptotic cells. The number of TUNEL-stained cells was expressed as RGCs per mm².

NGF Determination

Retinas from 24 animals (n=6 retinas per each experimental group) were homogenized by ultrasonication in extraction buffer (10 mM Tris-acetate, pH 7.4; 100 mM NaCl; 1 mM ethylenediamine-tetraacetic acid; 1 mM ethyleneglycol-tetraacetic acid; 1% Triton X-100; 10% glycerol; 0.1% SDS; 20 mM Na₄P₂O₇; 2 mM sodium-orthovanadate, Na₃VO₄; 1 mM NaF; 2 μ g/mL aprotinin; 1 mM phenylmethylsulfonyl fluoride; and 1 μ g/mL leupeptin), centrifuged at 4°C for 20 minutes at 12,000 rpm. The concentrations of NGF in the supernatants were determined by the use of an enzyme-linked immunosorbent assay kit following the instructions provided by the manufacturer (NGF Emax Immunoassay System; Promega, Madison, WI).

Statistical Analysis

All statistical evaluations were performed using the StatView package for Windows. The data are expressed as means \pm SEM. A *P* value of less than 0.05 was considered significant. Post-hoc comparison within logical sets of means was performed using Tukey test.

RESULTS

EIOP After Hypertonic Saline Injection

Figure 1 shows the globes with the episcleral vein (arrow; Fig. 1A) and the site of injection (Fig. 2B) of hypertonic saline with a glass microneedle. As shown in Figure 1C, injection of hypertonic saline leads to elevation of IOP, reaching greatest increase 7 days postinjection (42.7 ± 3.7 mm Hg as compared with 26.9 ± 2.6 mm Hg of CV), and remained elevated for the following 7 weeks. Ocular administration of NGF has no effect on IOP.

NGF and RGC Distribution in Glaucomatous Untreated and NGF-treated Eyes

Figure 2A reports the concentration of NGF in the retina with and without EIOP of NGF-treated and untreated rats. The level of NGF in the GLV retina decreased compared with baseline levels, at 7 weeks after induction of IOP (from 89 ± 8.5 to 62 ± 5.2 pg/g; *P* < 0.05). Ocular administration of NGF to glaucomatous eye (GLNGF) for 7 weeks enhanced retinal NGF levels (from 84 ± 8.0 to 151 ± 21.3 pg/g; *P* < 0.01). An increase in retinal NGF was also found in the control retina treated with NGF (CNGF; from 85 ± 10 to 139 ± 10.5 pg/g; *P* < 0.05).

Histologic analysis of the retina shows cell loss in the retinal ganglion layer after hypertonic saline injection: GLV (Fig. 2C) compared with CV (Fig. 2B). Ocular administration of NGF for 7 consecutive weeks prevented RGC loss in GLNGF (Fig. 2D). Quantitative analyses indicate that EIOP causes nearly 50% loss of the RGCs and that after NGF treatment, this loss is less than 20% (Fig. 2E).

The EIOP Causes Loss of NGF Receptors in RGCs

Under normal conditions, RGCs of rats express TrkA and p75 NGF receptors (Figs. 3A, H). Seven weeks after induction of EIOP, the number of TrkA-positive RGCs decreases (Fig. 3B vs. Fig. 3A) and this decrease is prevented by ocular administration of NGF (Fig. 3C vs. Fig. 3B). As reported in Figure 3E, these differences are statistically significant (GLV vs. CV: **P* < 0.01; GLNGF vs. GLV: ***P* < 0.05). Moreover, results of western blotting analysis, shown in Figures 3F and G, indicated that the TrkA protein expressed in the glaucomatous retina decreases and that NGF administration nearly prevented this decrease.

Figure 3 reports also the effect of EIOP on the expression of p75 receptor by RGCs. EIOP reduces the number of p75-positive cells (Fig. 3I vs. Fig. 3H) and this decrease is prevented by ocular NGF administration (Fig. 3L vs. Fig. 3I). As reported in Figure 3N, both the decrease of p75 expression in glaucomatous retina and the protective effect of NGF administration are statistically significant (GLV vs. CV: **P* < 0.01; GLNGF vs. GLV: ***P* < 0.05). Western blotting analysis, shown in Figures 3O and P, showed that the p75 protein present in the retina decreases in glaucomatous retina and that this decrease is nearly prevented by ocular NGF administration.

NGF Ocular Administration Prevents Apoptotic RGC Death

To assess whether the reduced RGCs number was due to cell death, we next investigated the expression of biomarkers involved in cell death and cell survival. The RGCs of rats with EIOP are markedly stained by TUNEL (arrows in Fig. 4B), a biomarker of cell death, and are

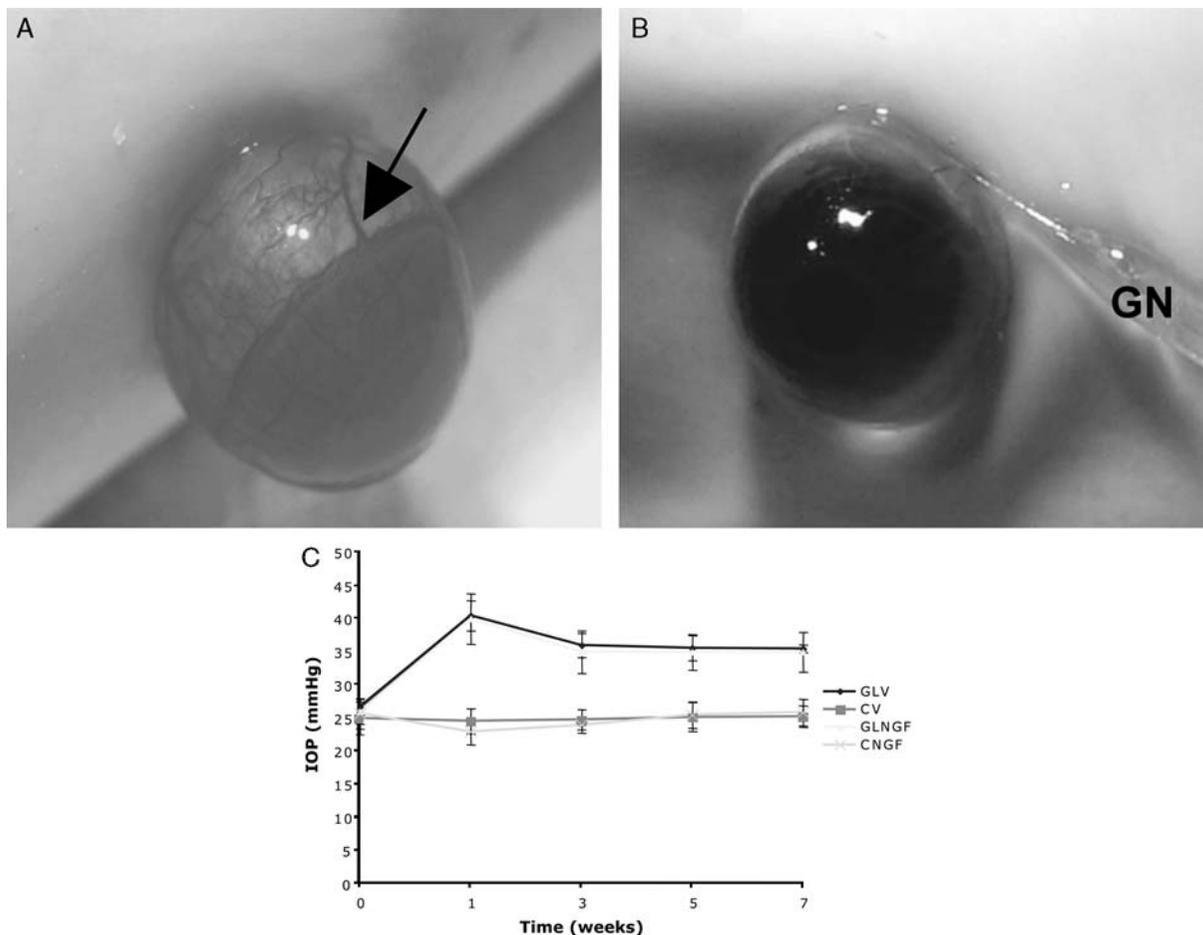


FIGURE 1. Representative microphotographs showing the episcleral vein (arrow) of rat eyes (A), the site of insertion of the glass microneedle (GN) in the episcleral vein (B), and the time-course IOP of rats injected and non-injected with hypertonic saline (C). As reported in this figure, the injection of hypertonic saline in the episcleral vein causes a significant elevation of IOP as compared with control, reaching peak after 1 week ($P < 0.01$). As indicated, nerve growth factor (NGF) eye administration is unable to normalize the IOP. CNGF indicates control eye treated with NGF; CV, control eye treated with vehicle solution; GLNGF, glaucomatous eye treated with NGF; GLV, glaucomatous eye treated with vehicle solution; IOP, intraocular pressure.

absent in control retina (Fig. 4A) and nearly absent in NGF-treated retinas (Fig. 4C). Quantitative analysis reported in Figure 4D revealed that the effect of NGF in preventing RGC apoptotic death is statistically significant ($P < 0.01$).

NGF Administration Reduces the Number of Degenerating ON Axons Induced by EIOP

Figure 5 shows representative electron microscopic sections of normal (Fig. 5A), glaucomatous (Fig. 5B), and NGF-treated glaucomatous (Fig. 5C) ON of a rat 7 weeks after induction of EIOP. As indicated by arrows, the ON of glaucomatous NGF-untreated rat shows degenerating axon profiles, characterized by varicosities or swelling, vacuolization, debris, and also signs of degenerating cells, most probably oligodendrocytes, nearly absent in the control and NGF treated axons. Quantitative determination reported in Figure 5D indicates that these differences are statistically significant ($P < 0.01$).

DISCUSSION

Using a rat model of GL induced by EIOP, we have investigated the role of NGF on damaged RGCs and ON axons. The results show that EIOP causes a transient local increase of NGF in the retina, followed by a significant decreases leading ultimately to RGC death through apoptotic mechanisms. We found that EIOP causes loss of RGCs, degeneration of ON axons, and a significant reduction of NGF, associated with a marked lower expression of low and high NGF-receptor by RGCs. EIOP causes a significant increase of TUNEL staining by RGCs, suggesting that degeneration of RGCs occurs through apoptotic mechanisms. These changes are markedly reduced by daily administration of eye NGF application, suggesting that NGF can protect the progressive degeneration of RGCs induced by EIOP.

Moreover, this study revealed that NGF delivered as eye drops protects RGCs and ON axons from degeneration and this noninvasive delivering method exerts its protective action in the absence of side effects. These observations support and extend earlier findings that NGF

A
Concentration of NGF protein (pg/g) in the retina of glaucomatous and non-glaucomatous rats under different experimental conditions.

Treatment	Weeks after treatment			
	0	1	3	7
CV	87±8.5	86±9.2	89±7.5	94±8.3
CNGF	85±10	117±11.9	116±10.4	139±10.5
GLV	89±8.5*	98±7.5*	104±7.0*	62±5.2**
GLNGF	84±8.0§	112±9.9§	137±12.7 §	151±21.3***

CV=non-glaucomatous eye treated with vehicle; CNGF=non-glaucomatous eye treated with NGF; GLV= glaucomatous eye treated with vehicle; GLNGF=glaucomatous eye treated with NGF. Data are the average ± S.E.M. of 5 samples in each experimental group. **p<0.01 compared to *; ***p<0.01 compared to §.

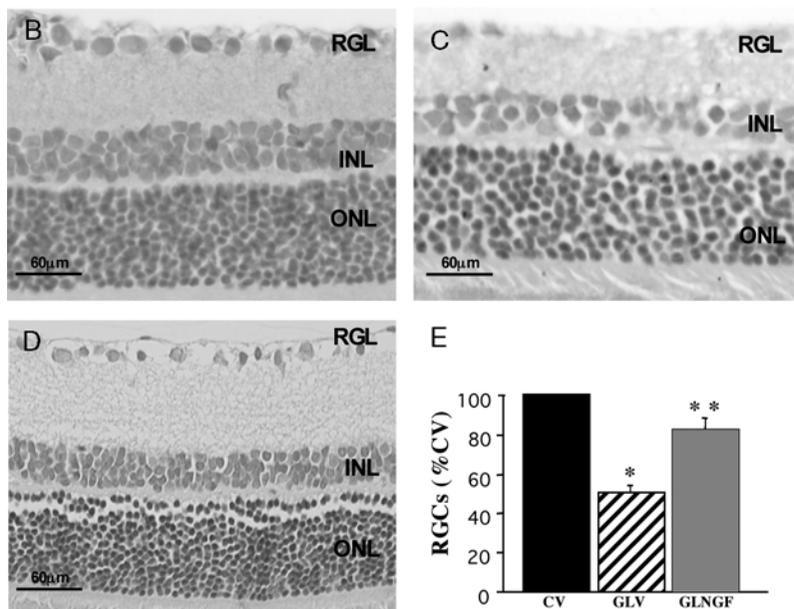


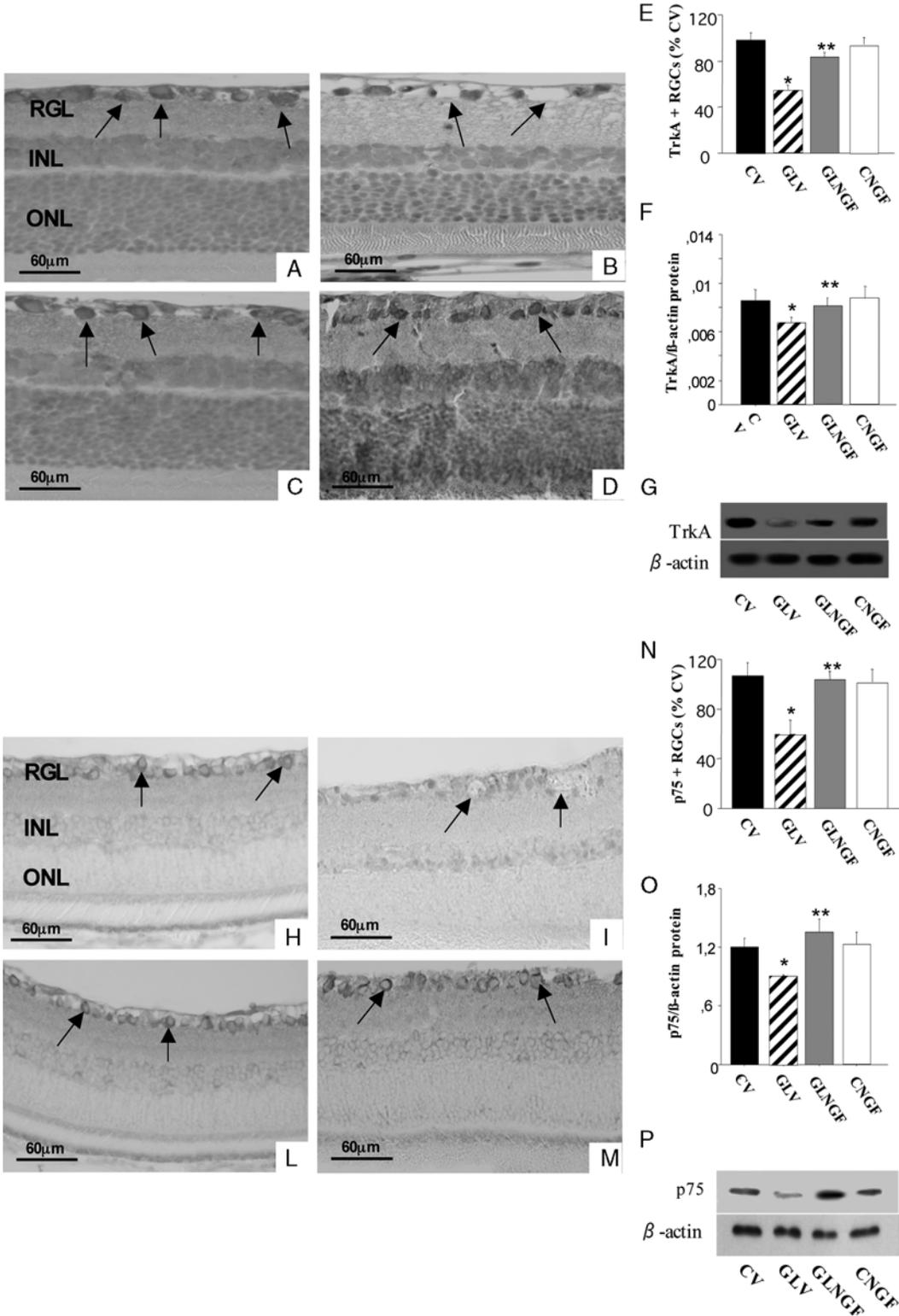
FIGURE 2. Time-course changes and statistic comparison between the end of treatment (7 wk) and 0 to 3 weeks reported in horizontal orientation. The concentration of NGF increases in the retina of glaucomatous retina (GLV) during the first 3 weeks (89±8.5, 98±7.3 vs. 104±7.0), thereafter the levels of NGF decreased significantly (62±5.2 vs. 104±7.0). In the glaucomatous retina treated with NGF (GLNGF), the level of NGF is elevated throughout the NGF treatment. An increase in NGF was also found in the retinas of normal rats treated with NGF (CNGF). Representative microscopic fields of normal (B), glaucomatous (C), and glaucomatous NGF-treated retinas (D), stained with hematoxylin-eosin showing the 3 retinal layers, the retinal ganglion layer (RGL), inner retinal layer (INL), and the outer retinal layer (ONL). Note that 7 weeks after hypertonic saline injection, the EIOP causes a marked decrease in the number of RGCs in the glaucomatous retina, compared with control retina. Quantitative evaluation (E) indicates that EIOP causes a statistically significant decrease in the number of RGCs compared with the control retina, whereas ocular administration of NGF largely prevented this decrease (*P<0.01 compared to CV, and **P<0.05 compared to GLV). B to D, Scale bars: 60 μm. EIOP indicates elevated intraocular pressure; NGF, nerve growth factor; RGCs, retinal ganglion cells.

and NGF receptors play a critical protective action on RGCs.^{15–17,20,21} More specifically, they indicate that shortage of NGF combined with abnormal expression of NGF-receptors are important key events that can lead to progressive RGC degeneration in GL and that NGF ocular administration can reduce or prevents these deleterious effects.

The critical role of NGF and its receptors in cell survival and cell death are supported by a number of other studies demonstrating that the biologic effect of NGF is mediated by competition between the low-affinity p75 receptor and the high-affinity TrkA receptor³¹ and the number of NGF-receptive RGCs can be critically dependent on such feedback loop. This dynamic mechanism

regarding the role of NGF and/or NGF-receptor interaction can occur also in diabetic NGF-responsive cells. For example, recent studies showed that NGF supplementation produces significant changes in NGF-receptor expression, leading to a 19-fold increase in TrkA/p75.^{32,33}

The role of NGF on retinal cells has been suggested by a number of other earlier and recent findings. Thus, intra-ocular administration of NGF has been shown to reduce rat RGC degeneration after ON lesion,¹⁴ in rabbit ocular hypertension,¹⁹ and in rats with inherited retinopathy.¹⁷ It has



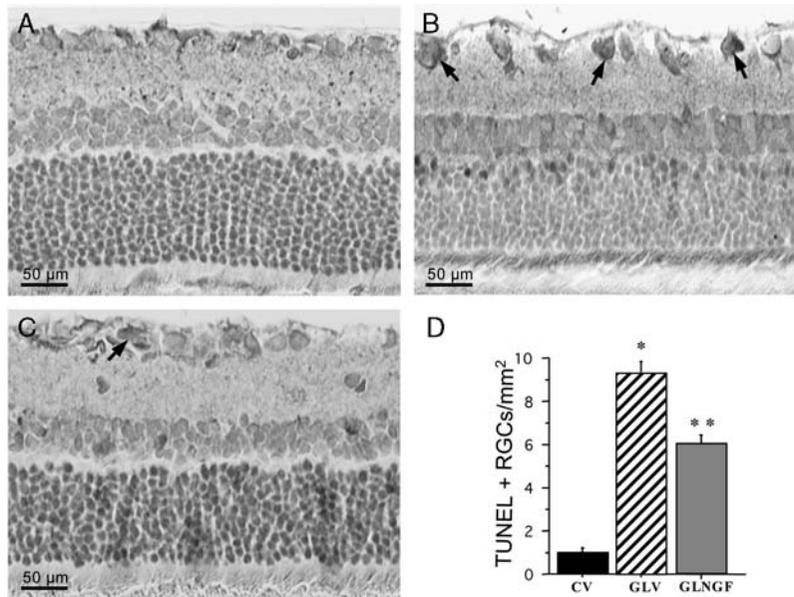


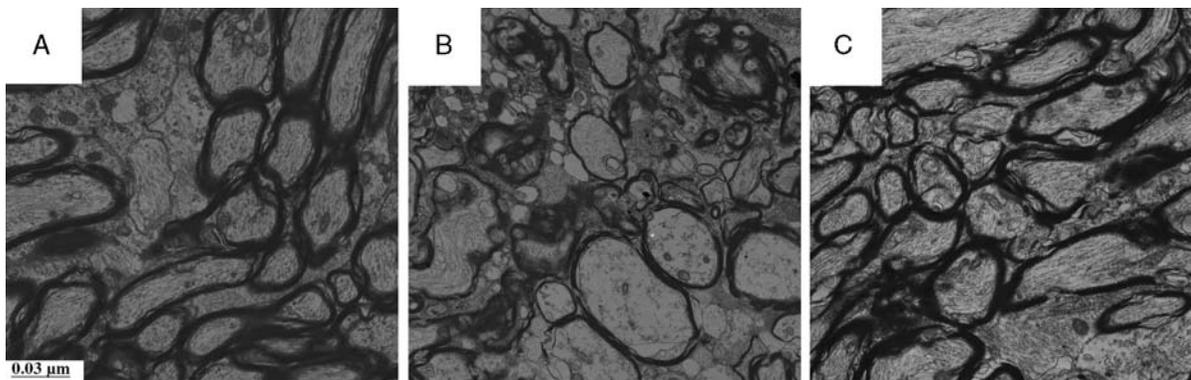
FIGURE 4. Representative retinal sections of control (A), glaucomatous (B), and glaucomatous eyes treated with NGF (C) stained by TUNEL, a marker for cell death. Note the presence of TUNEL-positive retinal ganglion cells (RGCs) (arrows) in glaucomatous eyes compared with control and glaucomatous eyes treated with NGF. TUNEL-positive cells in the retinal ganglion layer of glaucomatous eyes are indicated with arrows. These differences are statistically significant (D) [$*P < 0.01$ vs. control eye treated with vehicle solution (CV); $**P < 0.05$ vs. glaucomatous eye treated with vehicle solution (GLV)]. A to C, Scale bars: 50 μ m. GLNGF indicates glaucomatous retina treated with NGF; NGF, nerve growth factor; TUNEL, terminal deoxynucleotidyl transferase-mediated deoxyuridine triphosphate-biotin nick-end labeling.

been reported that administration of high molecular weight protein to the ocular surface can reach the retina and brain neurons through^{34,35} and that topical ocular administration of NGF can be a noninvasive approach to deliver NGF to the posterior portion of the eye and into the brain. These latter studies have demonstrated the protective role of eye NGF application on retinal cells not only in laboratory animals,^{36–38} but also in humans.³⁹ Indeed, in a recent observation in 3 patients with progressive visual-field defects and severe abnormalities in photopic electroretinogram and visual evoked potential responses and dysfunction of the innermost retinal layers, ocular NGF application improved mean visual field defects in 2 patients, and stabilized the defect in the third patient. Improvement of visual field

persisted 90 days after discontinuation of treatment, indicating that changes induced by NGF had a long-lasting effect.³⁹

Recently, studies by Rudzinski et al⁴⁰ and Shi et al⁴¹ performed in animal model of ocular hypertension or GL lead to the hypothesis that neither NGF nor antagonist of the proapoptotic p75 protect damaged RGC. Our results are somehow in contrast with their findings. Why our findings are different is not clear. We believe, based also on the available data that these differences can be because of: (a) the different animal model of GL we used; (b) the time and dose of NGF treatment; and (c) a more detailed structural and biochemical analysis or a combination of (a), (b), and (c). Moreover, it should be also taken into

FIGURE 3. Expression of TrkA and p75 receptors in retina of rats with EIOp treated and untreated with NGF. Representative microscopic fields of TrkA-immunostained control (A), glaucomatous (B), glaucomatous NGF-treated (C), and non-glaucomatous NGF-treated (D) in the RGL. These immunostained preparations indicate a reduced presence of TrkA-positive RGCs (arrows) in rats with EIOp (B), compared with controls (A) and that ocular NGF administration nearly prevented this reduction (C). Quantitative analysis (E) revealed that the decrease of TrkA-positive cells in glaucomatous retina and the effect on NGF are statistically significant ($*P < 0.01$ vs. CV; $**P < 0.05$ vs. GLV). Western blotting analysis of TrkA protein expressed in the glaucomatous retina treated and untreated with NGF, compared with their controls (F and G). The result indicates that TrkA protein decreases in glaucomatous retina and NGF administration in glaucomatous retina reduces this deficit ($*P < 0.01$ vs. CV; $**P < 0.05$ vs. GLV). Representative microscopic fields of p75-immunostained control (H), glaucomatous (I), glaucomatous NGF-treated (L), and non-glaucomatous NGF-treated (M) in the RGL. These immunostained preparations revealed a decreased expression of p75-positive RGCs (arrows) in rats with EIOp (I), compared with controls (H) and that ocular NGF administration nearly prevented this reduction (L). Quantitative analysis (N) revealed that the decrease of p75-positive cells in glaucomatous retina and the effect on NGF are statistically significant ($*P < 0.01$ vs. CV; $**P < 0.05$ vs. GLV). Western blotting analysis of p75 protein expressed in the glaucomatous retina treated and untreated with NGF, compared with their controls (O and P). The data obtained indicates that p75 protein decreases in glaucomatous retina and NGF administration in glaucomatous retina reduces this effect ($*P < 0.01$ vs. CV; $**P < 0.05$ vs. GLV). A to D and H to M scale bars: 0.03 μ m. CNGF indicates control eye treated with NGF; CV, control eye treated with vehicle solution; EIOp, elevated intraocular pressure; GLNGF, glaucomatous retina treated with NGF; GLV, glaucomatous eye treated with vehicle solution; INL, inner retinal layer; NGF, nerve growth factor; ONL, outer retinal layer; RGCs, retinal ganglion cells; RGL, retinal ganglion layer.



D
Axons of the optic nerve of control (CV), glaucomatous treated with vehicle (GLV) and glaucomatous treated with NGF (GLNGF) rats showing degenerative profiles, as revealed by ultrastructural analysis

type of treatment	ON section/rat	Area/section/μm ²	degenerating profile/area
CV	6	40000	15.0±7.3
GLV	6	40000	230.0±15.4**
GLNGF	6	40000	60.0±5.4*

**p<0.01 compared to CV and *p<0.05 to CV

FIGURE 5. Representative electron-microscopic sections of control (A), glaucomatous untreated (B), and glaucomatous NGF-treated ON (C), 7 weeks after the induction of elevated intraocular pressure. Note retinal ganglion cell axons in the ON of glaucomatous eye (B) showing varicosities, swellings, and vacuolization (arrows), compared with control ON (A). In the glaucomatous ON treated with NGF, these degenerating signs are greatly reduced or nearly absent (C). These differences are statistically significant (D) (*P<0.01 vs. CV; **P<0.01 vs. GLV). A to C scale bars: 0.03 μm. NGF indicates nerve growth factor; ON, optic nerve.

consideration that NGF has been shown to protect damaged RGCs after ON section,¹⁴ after ocular ischemia,¹⁶ ocular hypertension,⁴⁰ and inherited retinitis pigmentosa.¹⁷

A key question raised by our observations is the mechanism through which NGF administration protects RGCs without lowering the IOP. The evidence that patients with low IOP still experience visual loss, and others with GL and ON degeneration display normal IOP,^{4,5} suggests that the EIOP might not be the only mechanism through which RGCs degenerate. Thus, changes in the neuronal environment, such as a neurotransmitter imbalance, influx of calcium into the cells, formation of free radicals, and depletion of growth factors can take part in retinal degeneration and RGC damages induced by elevated IOP. This hypothesis is suggested by observation after systemic application of brimonidine,⁴² chondroitin sulphate proteoglycan,⁴³ or memantine, an N methyl-D-aspartate open channel blocker.⁴⁴ On the basis of the available data that RGCs are highly receptive to NGF, a reasonable hypothesis is that the ocular NGF application reaches the posterior segment of the eye bindings to retinal cells and renders more resistant RGCs to the damage induced by EIOP. Moreover, the possibility that NGF can exert its action through other growth factors, including brain-derived neurotrophic factor, cannot be excluded.^{18,25}

In summary, our data suggest that the reduced presence of NGF and the loss of NGF receptors in retinal cells is an important step in the progression of GL. The reduction of degenerative events in RGCs and ON after

ocular NGF application suggests that NGF represents a possible useful molecule for GL by protecting RGC death and ON axon loss.

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